



中华医学会
CHINESE MEDICAL ASSOCIATION



中华医学会神经外科学分会
CHINESE NEUROSURGICAL SOCIETY

RESEARCH

Open Access



SIRT3 protects against early brain injury following subarachnoid hemorrhage *via* promoting mitochondrial fusion in an AMPK dependent manner

Xun Wu, Jianing Luo, Haixiao Liu, Wenxing Cui, Dayun Feng and Yan Qu*

Abstract

Background: Subarachnoid hemorrhage (SAH), an acute cerebrovascular accident, features with its high death and disability rate. Sirtuin3 (SIRT3) is a NAD⁺ dependent deacetylase which mainly located in mitochondria. Reduced SIRT3 function was indicated to involve in many disorders of central nervous system. Herein, we aimed to explore the neuroprotective effects of SIRT3 on SAH and to furtherly explore the underlying mechanisms.

Methods: Adult C57BL/6 J male mice (8–10 weeks) were used to establish SAH models. The pharmacological agonist of SIRT3, Honokiol (HKL), was injected in an intraperitoneal manner (10 mg/kg) immediately after the operation. Brain edema and neurobehavioral score were assessed. Nissl staining and FJC staining were used to evaluate the extent of neuronal damage. The changes of mitochondria morphology were observed with transmission electron microscopy. Western blot was used for analyzing the protein level of SIRT3 and the downstream signaling molecules.

Result: SIRT3 was downregulated after SAH, and additional treatment of SIRT3 agonist HKL alleviated brain edema and neurobehavioral deficits after SAH. Additionally, electron microscopy showed that HKL significantly alleviated the morphological damage of mitochondria induced by SAH. Further studies showed that HKL could increase the level of mitochondrial fusion protein Mfn1 and Mfn2, thus maintaining (mitochondrial morphology), protecting mitochondrial function and promoting neural survival. While, additional Compound C (CC) treatment, a selective AMPK inhibitor, abolished these protective effects.

Conclusions: Activation of SIRT3 protects against SAH injury through improving mitochondrial fusion in an AMPK dependent manner.

Keywords: Subarachnoid hemorrhage, SIRT3, Honokiol, Mitochondrial fusion, AMPK, Mfn1, Mfn2

Background

Subarachnoid hemorrhage (SAH), an acute cerebrovascular accident, features with its high death and severe neurologic impairment [1, 2]. SAH can be induced by many events, including aneurysm rupture, hypertensive arteriosclerosis, severe trauma, arteriovenous malformation, or abnormal vascular network at skull base [3]. Recently, it was suggested that early brain injury (EBI) played a critical role in pathological process of SAH. EBI

refers to the direct brain damage before the occurrence of delayed vasospasm. Growing evidences showed that EBI might contribute largely to mortality and morbidity at the early period 24–72 h after the SAH [4]. The mechanisms of EBI are complicated, which include inflammatory reaction, oxidative stress injury, brain edema, and glutamate excitotoxicity [5].

Mitochondrial dysfunction was recognized as a critical pathological factor in SAH injury. Mitochondria are highly dynamic and undergo fusion and fission all the time under both physiological stimulation and pathological injuries. Normal mitochondrial morphology is

* Correspondence: yanqu0123@fmmu.edu.cn

Department of Neurosurgery, Tangdu Hospital, the Fourth Military Medical University, Xi'an 710038, Shaanxi, China



vital in the maintenance of mitochondrial function, energy metabolism, and calcium cycle and so on [6, 7]. Previous studies have confirmed that the damage of mitochondrial morphology contributed largely to the acute neurological injury [8, 9]. Mitofusins (Mfn1 and Mfn2), located onto outer membrane of mitochondria, play an indispensable role in regulating mitochondrial fusion. They participated in many cellular activities including oxidative stress, cell proliferation, cell apoptosis, and axonal transport of mitochondria [10–12]. However, the role of mitochondrial fusion in SAH has not been fully clarified.

Sirtuins are nicotinamide adenine dinucleotides (NAD)-dependent protein deacetylases [13]. SIRT3 is the most important deacetylase in mitochondria, which plays a central role in mitochondrial function and energy metabolism [14, 15]. SIRT3 was reported to protect against senile myocardial hypertrophy, tumors, aging-related diseases and so on [16, 17]. SIRT3 was also reported to be indispensable for the maintenance of mitochondrial dynamics in many diseases models. AMPK pathway participates in energy metabolism, helps to resist stress response, and promotes cell survival [18]. Previous studies have confirmed that there is a positive feedback loop between SIRT3 and AMPK, which protected ischemic brain damage [19]. Honokiol (HKL) is natural compound with a small molecular weight and is extracted from the bark of Magnoliaceae. HKL was demonstrated to be able to directly bind to sirt3, –enhances both protein levels and enzymatic activity at the same time, thereby alleviating the damage induced by cardiac hypertrophy [20]. In this experiment, we explored whether the activation of SIRT3 protected against SAH and investigated the underlying mechanisms focusing on SIRT3/AMPK pathway and mitochondrial fusion.

Methods

Animals and ethics

These experiments were under the permission of the Ethics Committee of the Fourth Military Medical University and were strictly conducted according to the guidelines of the National Institutes of Health Guide for the Care and Use of Laboratory Animals. Healthy C57BL/6J mice (weighed from 20 to 25 g, aged from 8 to 10 weeks, male) were purchased from the animal center of the Fourth Military Medical University. All mice were kept in animal facilities with specific pathogen-free conditions and fed well.

SAH surgery and treatments

In vivo subarachnoid hemorrhage model was established as described as below. Anesthetized mice with 2% pentobarbital sodium and kept them in a suitable position. The skin was cut and the tissue was separated to

uncover right common carotid artery bifurcation. Then after ligating and dissecting right external carotid artery, inserted the sharpened suture into the right internal carotid artery and pierced the artery approximately 10 to 12 mm inward from the common carotid artery bifurcation. And then insert 3 mm further for perforating the bifurcation of the anterior and middle cerebral arteries, staying about 20 s before withdrawal. Sham group mice were operated under the same experimental methods, except for the 3-mm insertion. HKL solution (10 mg/kg) was injected intraperitoneally immediately after the surgery.

Experiment design

Experiment 1: To explore the changes of SIRT3 in various time points after SAH, 56 mice (8 mice came from Sham group, and 48 mice came from SAH group which were survived from the initial total 63 mice) were randomly and equally assigned into seven groups (each contained 8 mice): Sham group, and six experimental groups at various time points, including 3 h, 6 h, 12 h, 24 h, 48 h, and 72 h after SAH. All experimental mice were sacrificed in indicated time points following the surgery to detect SIRT3 level to choose a suitable time point for the following experiments.

Experiment 2: Exploring the underlying mechanism of the protective effects of SIRT3 in vivo. Ninety-six mice (24 mice came from Sham group and 72 mice are survived from the initial total 103 mice) were randomly and equally assigned into the groups listed as follows (each contained 24 mice): Sham group, SAH group, SAH + HKL group, and SAH + CC group 24 h following SAH; 24 mice were randomly divided three parts: 8 mice were tested for behavioral disturbances and then brain water content measurement; 8 mice were used for western blot and transmission electron microscopy; the last 8 mice were collected for Fluoro-Jade C (F-JC) staining and Nissl staining.

Measurement of brain water content

Brain edema was assessed at 24 h after SAH. Mice brain was weighed once after the removal to record the wet weight, then put it in the oven with temperature from 95 to 100 °C for 72 h to get the dry weight. Brain water content was assessed and calculated through a computational formula: $(\text{wet weight} - \text{dry weight}) / \text{wet weight} \times 100\%$.

Modified neurological severity score

We take out the modified neurological severity score (mNSS) to assess the extent of neurological functional impairments 24 h after SAH as previous study described [21]. The mNSS which contains sensory, motor, and reflex tests was graded from 0 score to 18 score (0 score

refers to the normal score; 18 score refers to the maximal deficit score). These experiments were conducted by two observers who were blind to all groups.

Nissl staining

After anesthesia, mice were perfused with pentobarbital sodium; mice brains were removed and fixed with paraformaldehyde for approximately 4 h, following by being dehydrated in doses of 10%, 20%, and 30% sucrose in turn. Then brain tissue was embedded in paraffin and sliced into sections about 15 μm . Then, dewaxed and rehydrated the slices. Stained selected brain slices with 1% methyl violet (10–20 min), then treated with Nissl differentiation solution (4–8 s) and rinsed with purified water. The slices were then treated with gradient ethanol and xylene. Finally, observed and photographed the sections under an optical microscope.

FJC staining

The brains were removed, fixed, dehydrated, and sliced into pieces about 15 μm as described above. The slices were then incubated with 1% sodium hydroxide solution in 80% ethanol (5 min), then rehydrated in 70% ethanol and purified water (2 min) respectively. Slices were incubated with 0.06% potassium permanganate (10 min), washed with purified water (3 min), and incubated with 0.0001% F-JC solution (Millipore, Temecula, CA, USA 15 min). The slices were then washed three times with purified water for 1 min each time. The images were collected by A1 Si confocal microscope (Nikon, Japan), calculating the number of FJC positive neurons.

Transmission electron microscope

Mice were sacrificed and perfused at 24 h after SAH operation as described above, which was followed by perfusion of 0.9% saline and 4% paraformaldehyde, respectively. Mice were sacrificed and perfused at 24 h after SAH operation as described above. Brains were removed and tissues around hematoma were cut and trimmed into 1–2 mm wide blocks which were then fixed in 4% glutaraldehyde for overnight. The slices were then post-fixed with 1% osmium tetroxide (1 h). After that, they were dehydrated with ethanol immersion, which was followed by embedded in resin. Then, cut the pieces into 80-nm sections. Lastly, observed the ultrathin sections with a JEM-1400 electron microscope (JEOL, Tokyo, Japan) and captured the micrographs with a charge coupled device camera (Olympus, Tokyo, Japan).

Western blot

We separated protein samples with SDS-PAGE gels. After that, it was transferred to PVDF membranes (Millipore, USA). Then blocked the membranes with non-fat milk (5%) which was diluted with TBST, and

incubated the membranes with primary antibodies overnight at 4 °C. Second day, after three times 10-min TBST washes, we incubated the membranes with the appropriate secondary antibodies (1:5000) at 27 °C for 2 h. Lastly, after the three times of 5-min TBST washes, detect the protein bands with a BioRad imaging system (Bio-Rad, USA). The primary antibodies were listed as below: SIRT3(1:1000, C73E3, Cell Signaling), Ac-lys (1:1000,9441S, Cell Signaling), phospho-AMPK (1:1000, D4D6D, Cell Signaling), AMPK (1:1000, 2532S, Cell Signaling), Mfn2 (1:1000, D2D10, Cell Signaling), Mfn1 (1:1000, A9880, ABclonal), Bax (1:1000, gtx32465, Gene Tex), Bcl2 (1:1000, gtx100064, Gene Tex), β -actin(1:3000, wh096194, Wanleibio).

Statistical processing

All data were presented as mean \pm SD. GraphPad Prism 6.0 software (GraphPad, USA) was used for the following statistical analysis. Comparison between multiple groups used one-way analysis of variance (ANOVA), following with the Tukey post hoc test for the analysis. Comparison between two groups used Student's *t* test (unpaired, two-tailed) for the analysis. Differences with *P* values < 0.05 were identified as statistically significant.

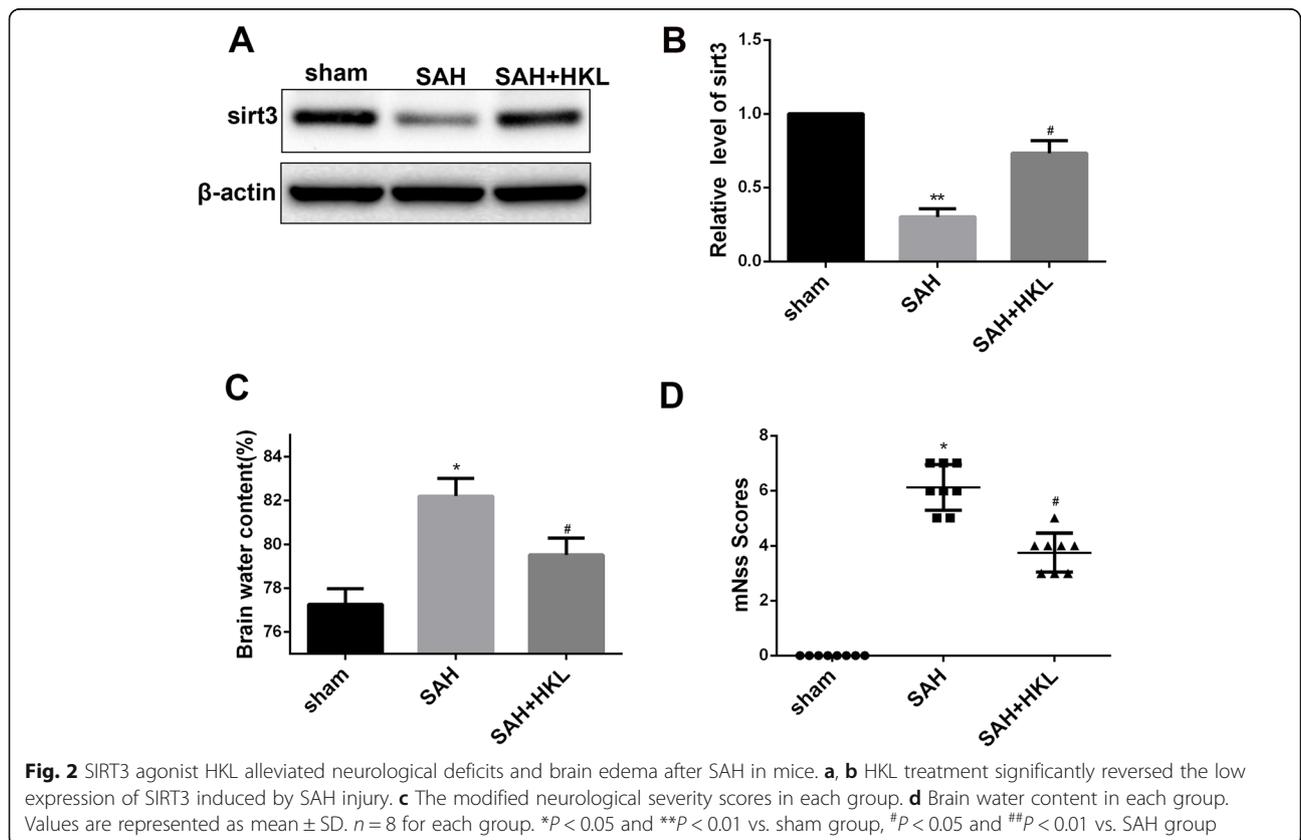
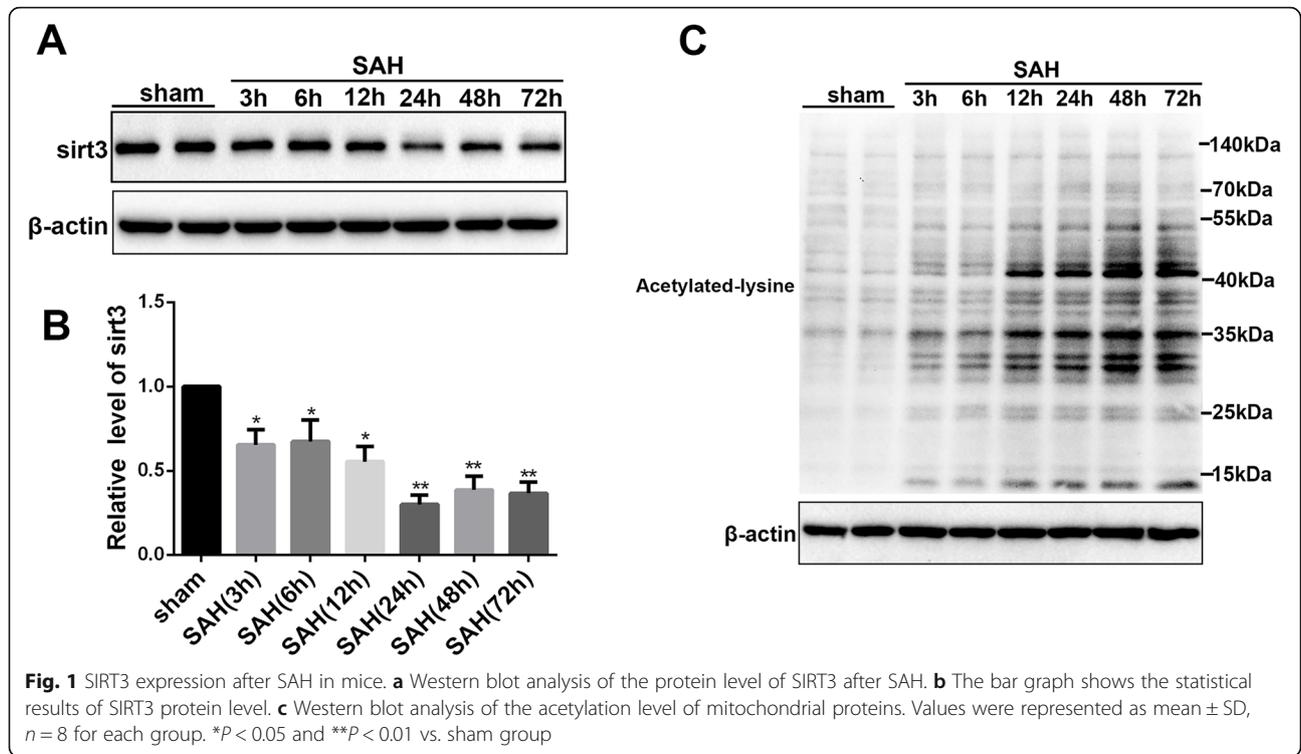
Results

SIRT3 expression after SAH injury in mice

In order to explore the time course of SIRT3-level in SAH, we detected the SIRT3 protein level at different time points including 0 (sham group), 3, 6, 12, 24, 48, and 72 h after SAH. The experiment revealed that the protein level of SIRT3 declined to the minimum at 24 h following SAH (Fig. 1a, b $P < 0.05$). SIRT3 is located in mitochondrial matrix and it plays a central role in regulating mitochondrial function via protein deacetylation to enhance their enzymatic activity. Disruption of SIRT3 activity contributed to the increased acetylation of mitochondrial proteins and impaired mitochondrial function. Then, we isolated the mitochondria to detect the acetylation level of mitochondrial proteins. Compared with sham group, the level of acetylation in SAH group was increased in a time-dependent manner (Fig. 1c, $P < 0.05$). These showed that the level of SIRT3 was decreased and the function of SIRT3 was impaired after SAH. So we take 24 h after SAH as the time point for the following experiments.

SIRT3 agonist HKL alleviated neurological function deficits and brain edema after SAH in mice

Western blot indicated that additional HKL treatment significantly reversed the low level of SIRT3 induced by SAH injury (Fig. 2a, b $P < 0.05$). Then we measured brain water content and took behavioral tests of mice to evaluate the effect of HKL on SAH. The



results revealed that brain water content increased significantly after SAH injury, which were significantly reduced by additional HKL treatment (Fig. 2c, $P < 0.05$). Similarly, we found significant neurological impairment emerged after SAH injury, while HKL treatment significantly improved the neurological function score after SAH (Fig. 2d, $P < 0.05$).

HKL ameliorated neuronal damage after SAH

Nissl staining and FJC staining were used to assess the effects of HKL on neural damage after SAH. Normal neurons feature with cytoplasmic Nissl staining. In contrast, loss of Nissl staining suggests the damaged neurons. The extent of neural loss was evaluated with the decreased number of Nissl-positive cells (Fig. 3a, c). The number of Nissl-positive cells was observed to be significantly downregulated in the SAH group (vs. sham group, $P < 0.05$), which was significantly reversed after the

treatment with HKL (vs. SAH group, $P < 0.05$). In addition, the protective effects were furtherly validated by FJC staining. FJC-positive cells represent degenerated neurons. We found that SAH insignificantly increased FJC-positive cells, which were reversed by additional HKL treatment (Fig. 3b, d, $P < 0.05$), indicating that HKL could alleviate neuronal damage after SAH.

HKL alleviated morphological damage of mitochondria after SAH

Electron microscopy was used to study the changes of mitochondrial morphology and structure. In the sham group, neurons were featured by long tubular mitochondrial structures, apparent cristae appearance, and some small globular mitochondrial structures (Fig. 4a, a1. Mitochondrial abnormalities were observed in neurons after SAH. In SAH group, amounts of small globular mitochondrial structures with mitochondrial fragments

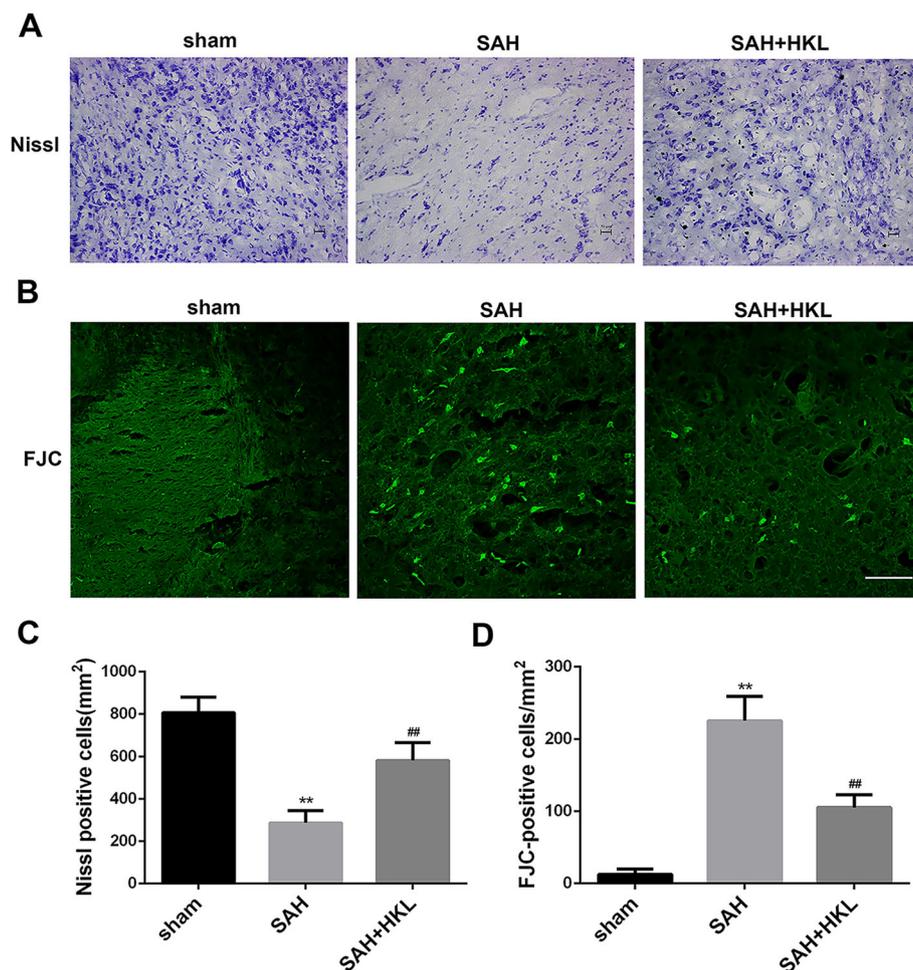
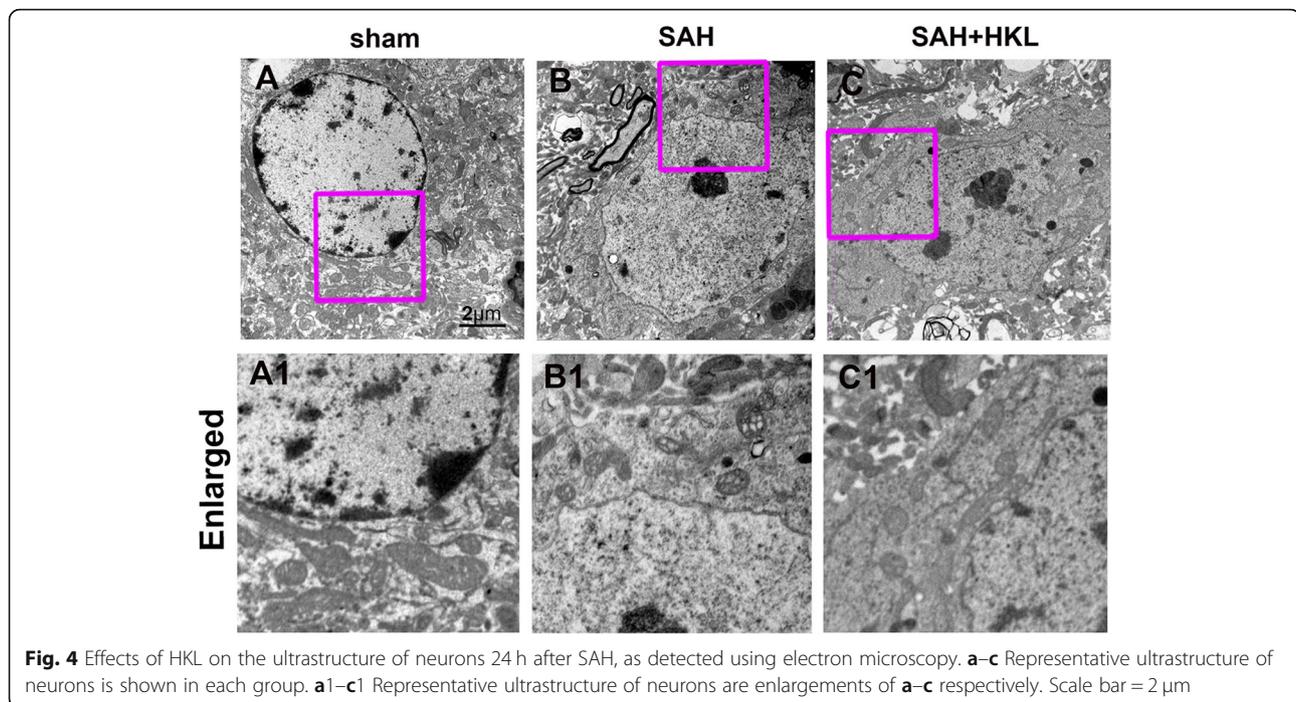


Fig. 3 HKL ameliorated neuronal damage after SAH in mice. **a** Representative images of Nissl staining and **c** quantitative analyses of Nissl positive neurons in each group 24 h after the operation are shown. Scale bar 50 μ m. **b** Representative images of FJC staining and **d** Quantitative analyses of FJC-positive cells in each group 24 h after the operation are shown. Scale bar 50 μ m. Values were represented as mean \pm SD, $n = 8$ for each group. * $P < 0.05$ and ** $P < 0.01$ vs. sham group, # $P < 0.05$ and ## $P < 0.01$ vs. SAH group



were observed. In addition, some mitochondria showed abnormal morphology after SAH, including swelling mitochondrial, ridge collapse, and membrane rupture (Figs. 4b, b1). Importantly, HKL therapy increased mitochondrial tubular networks, significantly reduced mitochondrial fragments, and largely maintained normal mitochondrial morphology (Fig. 4c, c1).

SIRT3 promoted mitochondrial fusion after SAH in an AMPK dependent manner

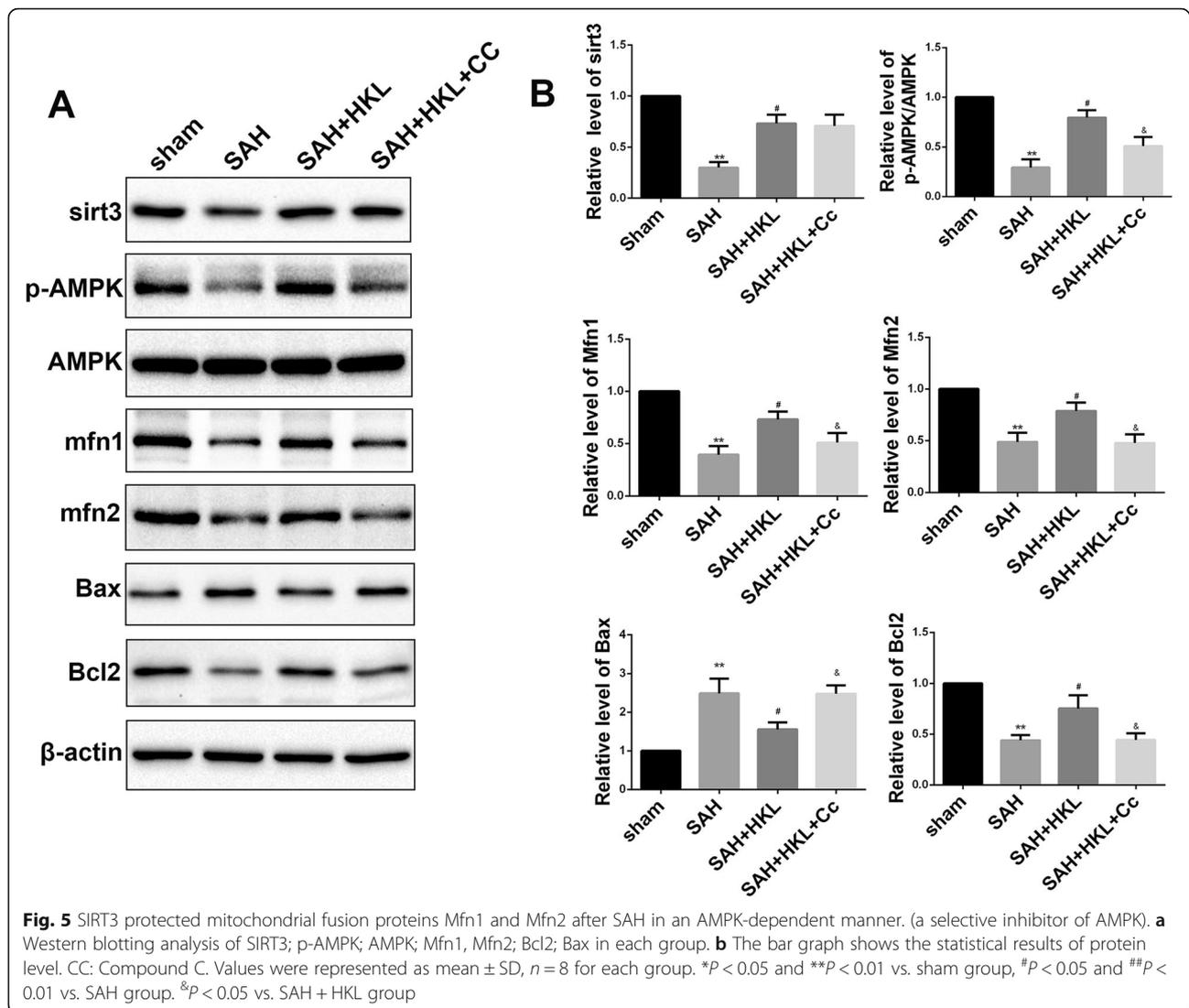
Western blotting results showed that the expressions of SIRT3, p-AMPK/AMPK, Mfn1, Mfn2, and Bcl2/Bax in SAH group were significantly declined than those in sham group (Fig. 5a, b, $P < 0.05$). However, the expression of SIRT3, p-AMPK/AMPK, Mfn1, Mfn2, and Bcl2/Bax significantly increased after HKL administration (Fig. 5a, b, $P < 0.05$), suggesting that HKL activated the SIRT3 and AMPK pathway, upregulated the expression of Mfn1 and Mfn2, protected mitochondrial morphology, and promoted neuron survival. CC treatment, a selective inhibitor of AMPK pathway, can largely counteract the protective effect of HKL (Fig. 5a, b, $P < 0.05$). It suggested that the protective effects of SIRT3 activator HKL on mitochondrial morphology were depended on the AMPK pathway.

Discussion

SAH is a life-threatening cerebrovascular emergency and features with high mortality and disability rates. Because the pathological process of SAH is complicated, the efficient intervention is still lacking. Mitochondrial dysfunction is identified as a major contributor to the early

brain injury of SAH [22, 23]. Herein, we explored the role of SIRT3, which is a NAD^+ -dependent deacetylase located in mitochondria, in SAH-induced brain injury. Previous researches indicated that SIRT3 protected against brain injury by inhibiting the activation of HIF1 α and ROS-mediated oxidative damage [24]. In this experiment, we found the protein level and deacetylation function of SIRT3 were significantly decreased after SAH. It is suggested that the impairment of SIRT3 may be one of the major factors contributing to the EBI of SAH. Then, after HKL treatment, brain edema, neurological dysfunction, and neuronal damage induced by SAH were significantly alleviated. It indicated that activation of SIRT3 could effectively protect SAH-induced brain injury.

Our further studies found that HKL significantly attenuated the morphological damage of mitochondria. Mitochondria constantly undergo fission and fusion, whose balance is indispensable for the maintenance of normal mitochondrial function. Mitofusins (Mfn1 and Mfn2) are of vital importance in regulating mitochondrial fusion. Previous studies showed that damaged mitochondrial oxidative phosphorylation and declined ATP production happened when Mfn2 was deleted [11, 12]. The mitofusins protein also—seemed to play additional roles in many process besides mitochondrial fusion. Mfn2 was found to take part in the axonal transport of mitochondria [25] and also was indicated in tethering endoplasmic reticulum (ER) and mitochondria [26]. Herein, we found HKL treatment can significantly elevate the level of Mfn1 and Mfn2, which may contribute the normalization of normal mitochondrial fusion.



AMPK pathway was demonstrated to play a critical role in regulating cellular energy metabolism, such as mitochondrial biogenesis, adaptive thermogenesis, glucose/fatty acid metabolism [18]. Previous studies implicated that there is a positive feedback loop between SIRT3 and AMPK [19]. Meanwhile, the activation of AMPK pathway was reported to promote the expression of Mfn1 and Mfn2 [8, 27]. In the current study, we found CC treatment, which is a selective inhibitor of AMPK pathway, counteracted the protective effect of HKL on mitochondrial fusion to a large extent. All these suggested that the protective effects of HKL on mitochondrial morphology relies on the activation of SIRT3/AMPK pathway.

There are several limitations have to be pointed out in this study. Firstly, we mainly discuss the protective effects of HKL and SIRT3 on EBI after SAH. Future studies

should further evaluate the long-term effect. Secondly, this study only focuses on the role of SIRT3 in regulating mitochondria fusion. We should also investigate the interaction of mitochondria with many organelles, such as endoplasmic reticulum, in the maintenance of normal physiological activity next time. Thirdly, although we have achieved satisfactory results in animal experiments, more studies including larger samples and more advanced technology applications are needed for the future clinical application.

Conclusion

Our study indicated that the activation of SIRT3 protected against SAH injury, and the underlying mechanism may be related to the regulation of mitochondrial fusion in an AMPK dependent manner.

Abbreviations

AMPK: Adenosine 5'-monophosphate (AMP)-activated protein kinase; F-JC: Fluoro-Jade C; HKL: Honokiol; mNSS: Modified neurological severity score; SAH: Subarachnoid hemorrhage; SIRT3: Sirtuin3

Acknowledgements

Thanks to Mr. Wu Hao who helped to provide constructive advice in the part of the discussion.

Authors' contributions

YQ conducted this project and designed the study. XW performed the experiments. J-NL and H-XL completed the statistics and interpreted the data. W-XC helped to draft the manuscript. D-YF revised the manuscript. All authors read and approved the final Manuscript.

Funding

This work was supported by grants from the State Key Program of National Natural Science Foundation of China 81630027 (to Dr. Qu), the National Natural Science Foundation of China 81571215 (to Dr. Qu), and the Chang Jiang Scholar Program of China (to Dr. Qu).

Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Ethics approval and consent to participate

All the animal experiments were carried out according to the Guide for the Care and Use of Laboratory Animals published by the U.S. National Institutes of Health (National Institutes of Health Publication No. 85-23, revised 1996) and was approved by the Ethics Committee of the Fourth Military Medical University (TDLL2017-03-190).

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Received: 25 September 2019 Accepted: 21 November 2019

Published online: 03 January 2020

References

- Zhao B, Rabinstein A, Murad MH, Lanzino G, Panni P, Brinjikji W. Surgical and endovascular treatment of poor-grade aneurysmal subarachnoid hemorrhage: a systematic review and meta-analysis. *J Neurosurg Sci.* 2017; 61(4):403–15.
- Nogueira AB, Nogueira AB, Esteves Veiga JC, Teixeira MJ. Multimodality monitoring, inflammation, and neuroregeneration in subarachnoid hemorrhage. *Neurosurgery.* 2014;75(6):678–89.
- Sehba FA, Hou J, Pluta RM, Zhang JH. The importance of early brain injury after subarachnoid hemorrhage. *Prog Neurobiol.* 2012;97(1):14–37.
- Helbok R, Schiefecker AJ, Beer R, Dietmann A, Antunes AP, Sohm F, et al. Early brain injury after aneurysmal subarachnoid hemorrhage: a multimodal neuromonitoring study. *Crit Care.* 2015;19:75.
- Echigo R, Shimohata N, Karatsu K, Yano F, Kayasuga-Kariya Y, Fujisawa A, et al. Trehalose treatment suppresses inflammation, oxidative stress, and vasospasm induced by experimental subarachnoid hemorrhage. *J Transl Med.* 2012;10:80.
- Roy S, Kim D, Sankaramoorthy A. Mitochondrial structural changes in the pathogenesis of diabetic Retinopathy. *J Clin Med.* 2019;8(9).
- Detmer SA, Chan DC. Functions and dysfunctions of mitochondrial dynamics. *Nat Rev Mol Cell Biol.* 2007;8(11):870–9.
- Dong H, Zhou W, Xin J, Shi H, Yao X, He Z, et al. Salvininin a moderates postischemic brain injury by preserving endothelial mitochondrial function via AMPK/Mfn2 activation. *Exp Neurol.* 2019;322:113045.
- Ren KD, Liu WN, Tian J, Zhang YY, Peng JJ, Zhang D, et al. Mitochondrial E3 ubiquitin ligase 1 promotes brain injury by disturbing mitochondrial dynamics in a rat model of ischemic stroke. *Eur J Pharmacol.* 2019;861: 172617.
- Zuchner S, Mersiyanova IV, Muglia M, Bissar-Tadmouri N, Rochelle J, Dadali EL, et al. Mutations in the mitochondrial GTPase mitofusin 2 cause Charcot-Marie-tooth neuropathy type 2A. *Nat Genet.* 2004;36(5):449–51.
- Rocha AG, Franco A, Krezel AM, Rumsey JM, Alberti JM, Knight WC, et al. MFN2 agonists reverse mitochondrial defects in preclinical models of Charcot-Marie-tooth disease type 2A. *Science.* 2018;360(6386):336–41.
- Nasrallah CM, Horvath TL. Mitochondrial dynamics in the central regulation of metabolism. *Nat Rev Endocrinol.* 2014;10(11):650–8.
- Onyango P, Celic I, McCaffery JM, Boeke JD, Feinberg AP. SIRT3, a human SIR2 homologue, is an NAD-dependent deacetylase localized to mitochondria. *Proc Natl Acad Sci U S A.* 2002;99(21):13653–8.
- Bell EL, Guarente L. The SirT3 divining rod points to oxidative stress. *Mol Cell.* 2011;42(5):561–8.
- Finkel T, Deng CX, Mostoslavsky R. Recent progress in the biology and physiology of sirtuins. *Nature.* 2009;460(7255):587–91.
- Albani D, Polito L, Forloni G. Sirtuins as novel targets for Alzheimer's disease and other neurodegenerative disorders: experimental and genetic evidence. *J Alzheimers Dis.* 2010;19(1):11–26.
- Polito L, Kehoe PG, Forloni G, Albani D. The molecular genetics of sirtuins: association with human longevity and age-related diseases. *Int J Mol Epidemiol Genet.* 2010;1(3):214–25.
- Carling D. AMPK signalling in health and disease. *Curr Opin Cell Biol.* 2017; 45:31–7.
- Duan WJ, Li YF, Liu FL, Deng J, Wu YP, Yuan WL, et al. A SIRT3/AMPK/ autophagy network orchestrates the protective effects of trans-resveratrol in stressed peritoneal macrophages and RAW 264.7 macrophages. *Free Radic Biol Med.* 2016;95:230–42.
- Pillai VB, Samant S, Sundaresan NR, Raghuraman H, Kim G, Bonner MY, et al. Honokiol blocks and reverses cardiac hypertrophy in mice by activating mitochondrial Sirt3. *Nat Commun.* 2015;6:6656.
- Chen J, Li Y, Wang L, Zhang Z, Lu D, Lu M, et al. Therapeutic benefit of intravenous administration of bone marrow stromal cells after cerebral ischemia in rats. *Stroke.* 2001;32(4):1005–11.
- Huang L, Wan J, Chen Y, Wang Z, Hui L, Li Y, et al. Inhibitory effects of p38 inhibitor against mitochondrial dysfunction in the early brain injury after subarachnoid hemorrhage in mice. *Brain Res.* 2013;1517:133–40.
- Zhang T, Wu P, Zhang JH, Li Y, Xu S, Wang C, et al. Docosahexaenoic acid alleviates oxidative stress-based apoptosis via improving mitochondrial dynamics in early brain injury after subarachnoid hemorrhage. *Cell Mol Neurobiol.* 2018;38(7):1413–23.
- Yang F, Zhou L, Wang D, Wang Z, Huang QY. Minocycline ameliorates hypoxia-induced blood-brain barrier damage by inhibition of HIF-1alpha through SIRT-3/PHD-2 degradation pathway. *Neuroscience.* 2015;304:250–9.
- Misko A, Jiang S, Wegorzewska I, Milbrandt J, Baloh RH. Mitofusin 2 is necessary for transport of axonal mitochondria and interacts with the Miro/Milton complex. *J Neurosci.* 2010;30(12):4232–40.
- de Brito OM, Scorrano L. Mitofusin 2 tethers endoplasmic reticulum to mitochondria. *Nature.* 2008;456(7222):605–10.
- Fan L, Wang J, Ma C. Pretreatment of bone mesenchymal stem cells with miR181-c facilitates craniofacial defect reconstruction via activating AMPK-Mfn1 signaling pathways. *J Recept Signal Transduct Res.* 2019;39(3):199–207.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

